-Original-

Plasma Immunoadsorption Therapy for Guillain-Barré Syndrome: Critical Day for Initiation

Hiroyuki Takei¹, Yuichi Komaba¹, Toshihiko Araki², Yasuhiko Iino¹ and Yasuo Katayama¹

¹Second Department of Internal Medicine, Nippon Medical School ²Department of Neurology, Kawaguchi Municipal Medical Center, Saitama

Abstract

Immunoadsorption plasmapheresis (IAPP) is a method of removing circulating immune factors that is used to treat Guillain-Barré syndrome (GBS). We retrospectively analyzed the data on our GBS patients. In 21 patients treated with IAPP, linear regression analysis showed that the time from the onset of symptoms to the initiation of IAPP was correlated with the time required for improvement by one Hughes functional grade. We investigated the critical day for initiating treatment, which we defined as the day when initiation of IAPP was significantly more likely to improve function by at least one Hughes grade when compared with the outcome in patients receiving supportive therapy (non-IAPP group). The critical day was found to be day 6 after the onset of GBS. (J Nippon Med Sch 2002; 69: 557–563)

Key words: immunoadsorption plasmapheresis, Guillain-Barré syndrome, timing of therapy, clinical improvement, regression analysis

Introduction

Guillain-Barré syndrome (GBS) is defined as an inflammatory polyradiculoneuropathy with a monophasic course that is characterized by rapidly progressive symmetrical muscle weakness and loss of tendon reflexes, followed by gradual remission. Pathological studies have suggested that this disease involves an autoimmune response directed against the peripheral nerves. Antibodies against gangliosides, which are cell surface components of nerves and other tissues, are often detected in patients with GBS, and an anti-ganglioside antibody has been demonstrated to cause motor nerve dysfunction in vitro¹. Campylobacter jejuni infection is one of the well-known antecedents of GBS. Interestingly, C. jejuni possesses a lipopolysaccharide component that structurally resembles GM1 ganglioside². Recent

studies have demonstrated that immunoadsorption plasmapheresis (IAPP) to remove autoantibodies is an effective treatment for GBS. The immunoadsorbents currently used are tryptophan or phenylalanine immobilized on polyvinyl alcohol gel. The latest time during the course of GBS at which initiation of IAPP can still be effective has not been clarified by previous studies, so we attempted to define this critical day in the present study.

Materials and Methods

A retrospective analysis was performed of the medical records and work histories of 38 patients with GBS. In all cases, the diagnosis of GBS was made by a neurologist whose evaluation included examination of the cerebrospinal fluid. The National Institute of Neurological and Communicative Disorders and Stroke (NINCDS) diagnostic criteria

Correspondence to Hiroyuki Takei, MD, Second Department of Internal Medicine, Nippon Medical School, 1–1–5 Sendagi, Bunkyo-ku, Tokyo 113–8603, Japan

E-mail: h-tokei@SC4.so-net.ne.jp

Journal Website (http://www.nms.ac.jp/jnms/)

for GBS were used.³ Patients with Fisher syndrome, Bell's palsy, Crow-Fukase syndrome, or a paraneoplastic syndrome were excluded. Five patients who underwent plasma exchange were also excluded (3 men and 2 women aged from 21 to 45 years). IAPP was performed in 21 GBS patients from 1985 to 1999 (IAPP group; 14 men and 7 women aged from 18 to 68 years). In all of these patients, IAPP was started within 2 days of admission. Although the duration from the onset of symptoms to initiation of IAPP tended to be longer in patients who were transferred to our hospital after a relatively long period, it was never deliberately delayed. Vascular access was obtained via a percutaneous transfemoral approach. During IAPP, plasma was separated from the cellular component using a membrane-type plasma separator (OP-05; Asahi Medical, Tokyo, Japan). The plasma was then passed through a TR 350 unit (Asahi Medical) containing a column of tryptophan immobilized on polyvinyl alcohol (PVA) gel to remove autoantibodies before it was returned to the blood. The volume of plasma processed during each IAPP session was 2 to 3 L, and the plasma flow rate was 18 to 25 mL/min. The mean number of IAPP sessions per patient was 6.3 ± 1.7 . No patient suffered from complications such as hypotension during IAPP or the development of a bleeding tendency, and no patient discontinued treatment prematurely.

Twelve untreated patients were admitted with GBS before 1984, when IAPP was not an established therapy (9 men and 3 women aged from 16 to 74 years) (non-IAPP group). All of the patients in both groups were classified functionally according to a grading scale proposed by Hughes et al (**Table 1**)⁴. There were no significant differences in sex, age, or clinical grade between the IAPP group and the non-IAPP group.

We defined improvement of GBS as a decrease of one grade on the Hughes scale. In the IAPP group the relationship between the time from the onset of GBS to the initiation of IAPP and the time until improvement by one grade was investigated by linear regression analysis using the least-squares method. A *p* value<0.05 was defined as indicating statistical significance. In the non-IAPP group, the mean \pm 1.96 SD of the time until improvement by one grade was also calculated. In the treated group, we used linear regression analysis to determine the critical days, which were defined as the longest time from the onset of symptoms to initiation of IAPP that still permitted improvement by one grade in less time than the mean minus 1.96 SD for the non-IAPP group (the lower limit of the 95% confidence interval). Patients receiving IAPP were then divided into early treatment and delayed treatment subgroups relative to the critical day, after which the time until improvement by one grade was compared between the early treatment subgroup, the delayed treatment subgroup, and the non-IAPP group by analysis of variance (ANOVA) with a post hoc Scheffe's test.

Results

Tables 1 and 2 list clinical information for the IAPP and non-IAPP groups. Among 21 patients recieving IAPP, one was classified as Hughes grade 1, 5 as grade 2, 3 as grade 3, 8 as grade 4, and 4 as grade 5. Among the 12 patients in the non-IAPP group, 4 were grade 2,4 were grade 3, and 4 were grade 4.

A significant correlation was observed between the interval from the onset of symptoms until the initiation of IAPP and the day until improvement by one grade (P < 0.05; r = 0.449) in the patients receiving IAPP (**Fig. 1**). The regression line was defined by the following equation: y = 0.923x + 11.920. In the non-IAPP group, the mean time ± 1.96 SD until improvement by one grade was 38.17 ± 20.71 days. According to regression analysis, the critical day was day 6 after the onset of GBS (Fig. 1). This means that symptomatic improvement of the patients treated with IAPP was faster than that of patients receiving supportive therapy alone when IAPP was started within 6 days of the onset. When the IAPP group was divided into patients treated within 6 days of the onset (early treatment subgroup) and those treated after more than 6 days

(delayed treatment subgroup), the day until improvement by one grade was significantly shorter in the early treatment subgroup than in the non-IAPP group or the delayed treatment subgroup, while the delayed treatment subgroup did not differ significantly from the non-IAPP group (**Fig. 2**).

Sex	Age	Sessions	Duration from onset to intiation of IAPP (days)	Clinical grade (Hughes)	Days needed to improve by one grade	Cranial nerve involvement	Sensory disturbance	Autonomic disturbance	Preceding event	Intial symptom
М	54	6	2	4	10		_	_	-	Muscle weakness of upper extremities
Μ	54	7	2	5	15	—	+	_	+	Muscle weakness of lower extremities
F	32	7	3	4	7	—	+	_	+	Numbness of four extremities
Μ	37	4	4	4	7	Dysphasia/Dysarthria	+	_	+	Muscle weakness of four extremities
F	25	9	4	3	8	—	+	_	+	Hypoesthesia of four extremities
F	26	3	4	3	8	—	+	_	+	Muscle weakness of lower extremities
Μ	48	7	8	2	19	—	+	_	+	Muscle weakness of lower extremities
М	46	7	3	5	21	Dysphasia	+	_	+	Dysphasia
М	43	5	5	3	12	—	_	_	+	Muscle weakness of four extremities
М	33	9	5	5	13	Dysphasia	_	_	_	Muscle weakness of four extremities
М	28	7	7	2	13	Facial palsy	_	_	+	Facial palsy
Μ	43	4	8	4	22	Facial palsy	+	_	-	Hypoesthesia of upper extremities
F	35	3	8	2	25	—	_	-	+	Muscle weakness of upper extremities
Μ	43	8	9	2	23	Dysarthria	_	_	-	Muscle weakness of lower extremities
F	58	7	9	4	45	—	+	_	-	Muscle weakness of lower extremities
М	68	5	17	4	79	—	_	_	_	Muscle weakness of four extremities
F	27	7	36	4	26	Facial palsy	_	-	-	Muscle weakness of four extremities
F	37	7	4	5	14	Facial palsy	+	-	+	Numbness of upper extremities
М	24	7	2	4	7	—	+	-	+	Numbness of lower extremities
М	25	7	12	1	14	Facial palsy/Dysarthria	_	_	_	Muscle weakness of lower extremities
Μ	18	7	20	2	21	—	_	-	+	Numbness of four extremities
	Sex M M F F M M M F F M M F M F M F M M F M M M F M M M M M F M M M F M M F M M F F M M M M M M F M	M 54 M 54 F 32 M 37 F 25 F 26 M 48 M 46 M 43 M 28 M 43 F 35 M 43 F 35 M 43 F 35 M 43 F 35 M 68 F 27 F 37 M 24 M 25	M 54 6 M 54 7 F 32 7 M 37 4 F 25 9 F 26 3 M 48 7 M 46 7 M 43 5 M 33 9 M 28 7 M 43 4 F 35 3 M 43 8 F 58 7 M 68 5 F 27 7 F 37 7 M 24 7 M 25 7	SexAgeSessionsonset to initiation of IAPPM5462M5472F3273M3744F2594F2634M4878M4673M3395M3395M2877M4389F5879M68517F27736F3774M2472M25712	SexAgeSessionsonset to initiation of IAPPClinical grade (Hughes)M 54 624M 54 725F 32 734M 37 444F 25 943F 26 343M 48 782M 46 735M 43 553M 33 955M 28 772M 43 484F 35 382M 43 892F 58 794F 27 7 36 4F 37 745M 24 724M 25 7121	SexAgeSessions onset to initiation of IAPP (days)Clinical grade (Hughes)improve by one gradeM 54 62410M 54 72515F 32 7347M 37 4447F 25 9438F 26 3438M 48 78219M 46 73521M 43 55312M 33 95513M 28 77213M 43 48422F 35 38225M 43 89223F 58 79445M 68 517479F 27 7 36 426F 37 74514M 24 7247M 25 712114	SexAgeSessionsonset to initiation of IAPP (days)Chinical grade (Hughes)improve by one gradeCranial herve involvementM5462410-M5472515-F327347-M374447Dysphasia/DysarthriaF259438-F263438-M4878219-M4673521DysphasiaM4355312-M3395513DysphasiaM4348422Facial palsyM4389223DysarthriaF5879445-M68517479-M68517479-M68517479-M427247-M25712114Facial palsy/Dysarthria	SexAge Sessions onset to initiation of IAPP (days)Clinical grade (Hughes)improve by one gradeCranial nerve involvementSensory disturbanceM5462410M5472515+F327347+M374447Dysphasia/Dysarthria+F259438+F263438+M4878219+M4673521Dysphasia+M4355312M3395513Dysphasia-M4348422Facial palsy-M4389223Dysarthria-M4389223Dysarthria-F5879445+M68517479F3774514Facial palsy+M247247-+M25712114Facial palsy/Dysarthria-	SexAge Sessions onset to initiation of IAPP (days)Clinical grade (Hughes)improve by one gradeClinical arefve involvementSensory disturbanceAutonomic disturbanceM5462410M5472515-+-F327347-+-M374447Dysphasia/Dysarthria+-F259438-+-M4878219-+-M4673521Dysphasia+-M4355312M3395513DysphasiaM4348422Facial palsyM4348422Facial palsyM4389223DysarthriaF5879445-+-M68517479M247247-+-M437214Facial palsyM43514Facial palsy <td>Sex Age Sessions of IAPP Chincal grade (Hughes) improve by one grade Cranial nerve involvement Sensory Autonomic disturbance Preceding event M 54 6 2 4 10 - - - - - + <t< td=""></t<></td>	Sex Age Sessions of IAPP Chincal grade (Hughes) improve by one grade Cranial nerve involvement Sensory Autonomic disturbance Preceding event M 54 6 2 4 10 - - - - - + <t< td=""></t<>

 Table 1
 Clinical features of the IAPP group

IAPP, immunoadsorption plasmapheresis.

Clinical grade (Hughes)

Grade 0; Healthy

Grade 1; Shiwing minor signs or symptoms of neuropathy but capable of manual work

Grade 2; Able to walk without support of a care but incapable of manual work

Grade 3; Able to walk with a care, appliance, or support

Grade 4; Conifined to bed or chairbound

Grade 5; Requiring assisted ventilation

Grade 6; Dead

Discussion

Immunological mechanisms are fundamental to the development of GBS, which is considered to be an autoimmune disease that targets the peripheral nerves. GBS often occurs between 1 and 6 weeks after a respiratory tract infection or gastroenteritis. which is frequently caused by Epstein-Barr virus, cytomegalovirus⁵, or C. jejuni⁶. Yuki² demonstrated that autoantibodies to GM1 ganglioside in serum from patients with GBS following C. jejuni infection react with an oligosaccharide protruding from the lipopolysaccharide core of the organism that is identical to the terminal tetrasaccharide of GM1 ganglioside, making this shared determinant a crossreactive antigen. Anti-GM1 ganglioside antibody is likely to cause peripheral nerve dysfunction, since it can bind with GM1, GD1b, and asialo GM17. Moreover, Takigawa⁸ showed that purified anti-GM1 antibodies block sodium channels at the nodes of Ranvier in the presence of complement. The GM1 epitope is mainly present on the nodes of Ranvier, the presynaptic membranes, and the motor neurons of the spinal cord^{9,10}. Patients who develop GBS following mycoplasma infection frequently have serum anti-galactocerebroside antibodies11, while patients developing GBS after cytomegalovirus infection often have anti-GM2 antibodies¹². Moreover, serum from patients with GBS often contains antibodies directed against various gangliosides, including GD1a, GD1b, GT1b, LM1, and asialo GM1 as well as GM1 or GM2¹³⁻¹⁶.

After the treatment of GBS with prednisolone was first attempted by Shy17, prednisolone was frequently used to treat this disease. However, Hughes subsequently found no significant difference between GBS patients treated with prednisolone and an untreated control group⁴, later extending these observations to high-dose intravenous methylprednisolone therapy with a similarly negative result¹⁸. Brettle¹⁹ was the first investigator to report on the use of plasma exchange for GBS. The French Cooperative Group then demonstrated the efficacy of plasma exchange for GBS in a large-scale controlled trial²⁰. Plasma exchange is currently recognized as effective for GBS and is often used as the initial therapy^{21,22}. However, plasma exchange may have

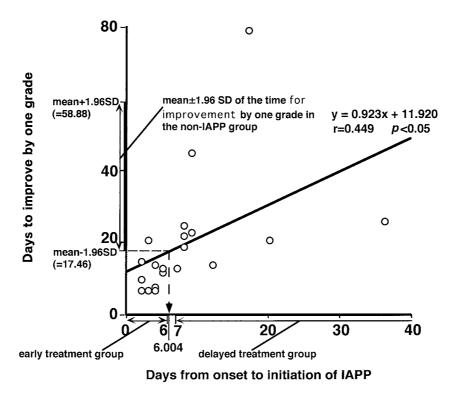


Fig. 1 Scatter plot of the time from the onset of symptoms to the initiation of plasmapheresis versus the time required for clinical improvement by one Hughes grade. A linear correlation is evident between the time until treatment and the time required for improvement by one grade. Data of case 5 overlays that of case 6. The regression line was defined by the following equation: y = 0.923 x + 11.920. Using this equation, improvement by one grade in the IAPP group occurred significantly faster than the mean minus 1.96 (the lower limit of the 95% confidence interval) of the time for the same improvement in the non-IAAP group, when IAPP was started within 6.004 days. Therefore, the critical day was day 6 from the onset of GBS.

numerous adverse effects, including allergic reactions or the transmission of infections such as hepatitis or acquired immunodeficiency syndrome when using fresh frozen plasma, and this therapy requires large volumes of replacement fluid. If albumin is used instead of fresh frozen plasma, the risk of transmitting infections during plasma exchange is probably reduced.

IAPP is an alternative method for removing circulating factors, such as IgG, IgM, IgA, and complement components², which does not require the used of a replacement fluid (such as fresh frozen plasma or albumin in saline) because the separated plasma is returned to the blood after removal of circulating factors.

We chose a PVA gel column containing tryptophan rather than one containing phenylalanine. While both kinds of column can selectively adsorb autoantibodies or immune complexes, Yuki found that the tryptophan column was more effective for adsorbing anti-ganglioside antibodies²³, probably because of the increased hydrophobicity contributed by the side chains of tryptophan.

The present study showed that IAPP should be initiated within 6 days of the onset of GBS. The GBS study group found that clinical improvement of patients treated with plasmapheresis was significantly faster than that of patients given supportive therapy alone when treatment was started within 7 days of the first symptoms. Although the reason why 7 days was the cut-off time was not discussed, this finding is similar to the present results²⁴. Tagawa demonstrated that more immunoglobulins and C3 were removed by plasma exchange than by IAPP using a tryptophanimmobilized polyvinyl alcohol gel column²⁵. Therefore, there is a possibility that the critical period might be extended by using plasma exchange instead of IAPP. We defined the critical day for initiating IAPP based on the premise that the time

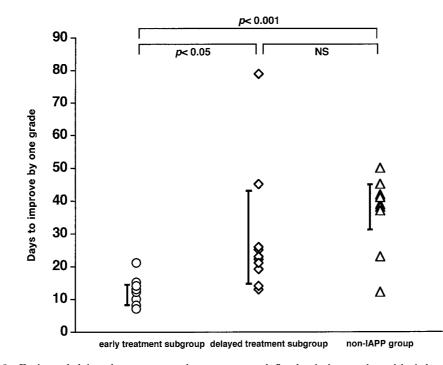


Fig. 2 Early and delayed treatment subgroups were defined relative to the critical day (day 6). Each scatter plot represents the time required for one Hughes grade of clinical improvement. Each bar represents the mean±SD of the time for improvement by one grade. Improvement was slower in the delayed treatment subgroup and the non-IAPP group than in the early treatment subgroup. Columns represent the mean values and bars show the standard deviation. NS, not significant.

for improvement by one grade had to be shorter than the 95% confidence interval (mean ± 1.96 SD) in untreated patients to provide a clear and consistent benefit. During the development of GBS, anti-GM 1 antibodies block sodium channels at the nodes of Ranvier in the early stage⁸, and damage to peripheral nerves occurs subsequently. It is desirable to prevent the progression of GBS before such pathologic changes occur, because it takes a certain amount of time to recover from these changes. The fact that the critical day for initiating IAPP is 6 days after the onset of GBS may reflect the occurrence of pathologic changes in the nurves. Birchem showed by electron microscopy that acutephase serum from GBS patients was cytolytic for myelin-related Schwann cells and could damage peripheral myelin in an experimental setting free of leukocytes or mononuclear cells²⁶. These findings suggest that antibodies and complement may play an important role in the acute pathology of GBS. Pathologically, GBS is categorized into acute demyelinating polyradiculopathy (AIDP), acute motor axonal neuropathy (AMAN), and acute motorsensory axonal neuropathy (AMSAN). In AIDP, Haymaker and Kernohan detected edema during the initial 3 to 4 days, followed by the onset of swelling and irregularity of the myelin sheath and axis cylinders on day 5, the appearance of a few lymphocytes on day 9, and phagocytosis on day 11²⁷. In the acute axonal pattern of GBS, the pathologic features are wallerian-like degeneration of fibers with little demyelination²⁸⁻³⁰.

Although there are relatively few macrophages in the spinal roots on day 7, endoneurial macrophages become numerous and foamy by day 18 in patients with this type of GBS. IAPP seems to prevent immunoglobulins and complement from causing direct damage to myelin sheaths or axons. There is also a possibility that IAPP may prevent lymphocyte cytotoxicity mediated by antigen-antibody reactions, or may prevent macrophages from binding immunoglobulins by removing antibodies.

References

 Arasaki K, Kusunoki S, Kudo N, Kanazawa I: Acute conduction block in vitro following exposure to antiganglioside sera. Muscle Nerve 1993; 16: 587–593. J Nippon Med Sch 2002; 69(6)

- Yuki N, Taki T, Inagaki F, Kasama T, Takahashi M, Saito K, Handa S, Miyatake T: A bacterium lipopolysaccharide that elicits Guillain-Barré syndrome has a GM 1 ganglioside-like structure. J Exp Med 1993; 78: 1771–1775.
- Asbury AK, Arnason BG, Karp HR, McFarlin DE: Criteria for diagnosis of Guillain-Barré syndrome. Ann Neurol 1978; 3:565–567.
- Hughes RAC, Newsom-Davis JM, Perkin GD, Pierce JM: Controlled trial of prednisolone in acute polyneuropathy. Lancet 1978; 2:750–753.
- Dowling PC, Bosch VV, Cook SD, Chmel H: Serum immunoglobulins in Guillain-Barré syndrome. J Neurol Sci 1982; 57: 435–440.
- Kaldor J, Speed BR: Guillain-Barré syndrome and *Campylobacter* jejuni: a serological study. Br Med J 1984; 288: 1867–1870.
- Yuki N, Yoshino H, Sato S, Miyatake T: Acute axonal polyneuropathy associated with anti-GM 1 antibodies following *Campylobacter* enteritis. Neurology 1990; 40: 1900–1902.
- Takigawa T, Yasuda H, Kikkawa R, Shigeta Y, Saida T, Kitasato H: Antibodies against GM1 ganglioside affect K + and Na + currents in isolated rat myelinated nerve fibers. Ann Neurol 1995; 37: 436–442.
- Corbo M, Quattrini A, Lugaresi A, Santoro M, Latov N, Hays AP: Patterns of reactivity of human anti-GM 1 antibodies with spinal cord and motor neurons. Ann Neurol 1992; 32: 487–493.
- Corbo M, Quattrini A, Latov N, Hays AP: Localization of GM1 and Gal (β 1–3) GalNAc antigenic determinants in peripheral nerve. Neurology 1993; 43: 809–814.
- Kusunoki S, Chiba A, Hitoshi S, Takizawa H, Kanazawa I: Anti-Gal-C antibody in autoimmune neuropathies subsequent to mycoplasma infection. Muscle Nerve 1995; 18: 409–413.
- Irie S, Saito T, Nakamura K, Kanazawa N, Ogino M, Nukazawa T, Ito H, Tamai Y, Kowa H: Association of anti-GM2 antibodies in Guillain-Barré syndrome with acute cytomegalovirus infection. J Neuroimmunol 1996; 68: 19–26.
- Fredman P, Vedeler CA, Nyland H, Aarli JA, Svennerholm L: Antibodies in sera from patients with inflammatory demyelinating polyradiculoneuropathy react with ganglioside LM1 and sulphatide of peripheral nerve myelin. J Neurol 1991; 238: 75–79.
- Gregson NA, Koblar S, Hughes RA: Antibodies to gangliosides in Guillain-Barré syndrome: specificity and relationship to clinical features. Q J Med 1993; 86: 111–117.
- Kusunoki S, Chiba A, Tai T, Kanazawa I: Localization of GM1 and GD1b antigens in the human peripheral nervous system. Muscle Nerve 1993; 16: 752–756.
- 16. Yuki N, Yoshino H, Sato S, Shinozawa K, Miyatake T: Severe acute axonal form of Guillain-Barré syndrome associated with IgG anti-GD1a antibodies. Muscle Nerve 1992; 15: 899–903.
- 17. Shy GM, McEachern D: Further studies of the

effects of cortisone and ACTH on neurological disorders. Brain 1951; 74: 354–362.

- Hughes RAC, for the Guillain-Barré syndrome steroid trial group. Ineffectiveness of high-dose of intravenous methylpredonisolone in Guillain-Barré syndrome. Lancet 1991; 2:1142.
- 19. Brettle RP, Gross M, Legg NJ, Lockwood M, Pallis C: Treatment of acute polyneuropathy by plasma exchange. Lancet 1978; 2: 1100.
- The French Cooperative Group on Plasma Exchange in Guillain-Barré syndrome: Appropriate number of plasma exchanges in Guillain-Barré syndrome. Ann Neurol 1997; 41: 298–306.
- French Cooperative Group on Plasma Exchange in Guillain-Barré Syndrome:Efficiency of Plasma Exchange in Guillain-Barré syndrome:role of replacement fluids. Ann Neurol 1987; 22: 753–761.
- 22. French Cooperative Group on Plasma Exchange in Guillain-Barré Syndrome: Plasma exchange in Guillain-Barré syndrome: one-year follow-up. Ann Neurol 1992; 32: 94–97.
- Yuki N: Tryptophan-immobilized column adsorbs immunoglobulin G anti-GQ1b antibody from Fisher's syndrome: A new approach to treatment. Neurology 1996; 46: 1644–1651.
- The Guillain-Barr é syndrome study group: Plasmapheresis and acute Guillain-Barré syndrome. Neurology 1985; 35: 1096–1107.
- Tagawa Y, Yuki N, Hirata K: Ability to remove immunoglobulins and anti-ganglioside antibodies by plasma exchange, double-filtration plasmapheresis and immunoadsorption. J Neurol Sci 1998; 157: 90–95.
- Birchem R, Mithen FA, L'Empereur KM, Wessels MM: Ultrastructural effects of Guillain-Barré serum in cultures containing only rat Schwann cells and dorsal root ganglion neurons. Brain Res 1987; 421: 173–185.
- Haymaker W, Kernohan JW: The Landry-Guillain-Barré syndrome. A clinicopathologic report of fifty fatal cases and a critique of the literature. Medicine 1949; 28: 59–141.
- Griffin JW, Li CY, Ho TW, Tian M, Gao CY, Xue P, Mishu B, Cornblath DR, Macko C, McKhann GM, Asbury AK: Pathology of the motor-sensory axonal Guillain-Barré syndrome. Ann Neurol 1996; 39: 17–28.
- Griffin JW, Li CY, Ho TW, Xue P, Macko C, Gao CY, Yang C, Tian M, Mishu B, Cornblath DR: Guillain-Barré syndrome in northern China. The spectrum of neuropathological changes in clinically defined cases. Brain 1995; 118: 577–595.
- Hafer-Macko C, Hsieh ST, Li CY, Ho TW, Sheikh K, Cornblath DR, McKhann GM, Asbury AK, Griffin JW: Acute motor axonal neuropathy: an antibodymediated attack on axolemma. Ann Neurol 1996; 40: 635–644.

(Received, January 7, 2002) (Accepted, July 18, 2002)